

Endocrine Disruptors and Human Health

G. Latini^{*1,2}, G. Knipp³, A. Mantovani⁴, M.L. Marcovecchio⁵, F. Chiarelli⁵ and O. Söder⁶

¹Division of Neonatology, Perrino Hospital, Brindisi, Italy; ²Clinical Physiology Institute, National Research Council of Italy (IFC-CNR); ³Purdue University, Department of Industrial and Physical Pharmacy, 575 Stadium Mall Dr., West Lafayette, IN 47907; ⁴Food and Veterinary Toxicology Unit, Dept. Veterinary Public Health and Food Safety, Istituto Superiore di Sanità, Viale Regina Elena, 299 00161, Rome; ⁵Department of Pediatrics, University of Chieti, via dei Vestini 5, Chieti, Italy; ⁶Department of Women's and Children's Health, Paediatric Endocrinology Unit, Karolinska Institute and University Hospital, S-17176, Stockholm, Sweden

Abstract: Endocrine-disrupting chemicals (EDCs) are a group of diversely natural compounds or synthetic chemicals that can interfere with the programming of normal endocrine-signalling pathways during pre- and neonatal life, thus leading to adverse consequences later in life. In addition, early life exposure to EDCs may alter gene expression and consequently transmit these effects to future generations.

Keywords: Endocrine-disruptors, environment, endocrine system, phthalates, pregnancy, neonate, fetal.

INTRODUCTION

Endocrine-disrupting chemicals (EDCs) are a large and increasing group of diversely natural compounds or synthetic chemicals present in the environment that include persistent halogenated pollutants, such as polychlorinated biphenyls (PCBs), polybrominated diphenylethers (PBDEs) and metabolites, industrial compounds, such as bisphenol A (BPA), alkylphenols and phthalate acid esters, as well as pharmaceuticals, pesticides, such as chlorpyrifos, fungicides including vinclozalin and phytoestrogens.

Man-made EDCs range across all continents and oceans. EDCs, which are typically present as complex mixtures and not as single substances, may mimic, block or modulate the synthesis, release, transport, binding, metabolism and/or elimination of natural endogenous hormones in wild animals and humans [1]. In particular, EDC may interfere with hormonal signalling systems and alter feedback loops in the brain, pituitary, gonads, thyroid, and other components of the endocrine system.

Growing evidence shows that EDC may also modulate the activity/expression of steroidogenic enzymes and steroidogenic pathways [2-5].

In addition, EDC can also promote activation of metabolic sensors, such as the peroxisome proliferator-activated receptors (PPARs) [6]. As a consequence, there is an increasing concern worldwide on the potential adverse effects of ED on human health, although their impact on human beings' health is not yet clear.

However, endocrine signalling pathways play an important role during prenatal differentiation; thus, developing organisms may be particularly sensitive to ED effects. In

fact, scientific evidence indicate that exposure to ED during critical periods of development can induce permanent changes in several organs, including molecular alterations, although the consequences of this disruption may not appear until later [7-11]. The mechanisms by which ED exert their action remain largely unclear; however, many ways have been identified by which ED can affect signal transduction systems [12].

Early life exposures to EDCs may alter gene expression *via* non-genomic, epigenetic mechanisms, including DNA methylation and histone acetylation, thus interfering with the germ-line. By contaminating the environment with ED human race might be permanently affecting the health of subsequent generations [13-15]. Within the broad ED topic we have focussed on specific issues, selected since they are highly relevant to the up-to-date assessment of potential human health risks from ED exposure.

ED IN THE FOOD CHAIN: HOW THEY INTERACT WITH NATURAL COMPOUNDS?

Diet is a significant source of exposure to ED for the general population, as well as a source of concern for consumers' health. One major issue is the "cocktail" effect: one cannot rule out additivity of different ED present in whole diet at low level, but hitting the same targets, e.g. nuclear receptors [16]. Furthermore, it is not just the daily dose that matters. Many ED can bioaccumulate in lipid fraction of tissues, originating a mixture "body burden" of contaminants of different origin that can include dioxins, polychlorinated biphenyls, chlorinated pesticides and their metabolites, as well as brominated flame retardants [17]. Other compounds may also concentrate in food chains, thus adding to the overall ED burden, e.g., organotins [18]. However, the modern conception of food toxicology cannot consider diet just as an exposure source of external harmful substances. Contaminants such as ED may interact with the same metabolic pathways as natural food components such as polyunsatu-

*Address correspondence to this author at the Division of Neonatology, Ospedale A. Perrino, s.s. 7 per Mesagne, 72100 Brindisi, Italy; Tel: +39-0831-537471; Fax: +39-0831-537861; E-mail:gilatini@tin.it

rated fatty acids, trace elements, vitamins and other bioactive substances (e.g. polyphenols) that cannot be considered nutrients as there is no recognized deficiency [19]. Dietary habits are related to socioeconomic status, cultural and religious factors, individual choices (e.g. vegetarianism/veganism); and dietary habits themselves may have the most important impact on the intake of both nutrients and contaminants. For instance, greater exposure to persistent ED is associated with the high consumption of fatty foods of animal origin [20, 21]. Thus, for specific food commodities a balanced evaluation is needed about contaminant-associated risks and nutritional benefits. A relevant example is represented by salmonids and other seafood, a useful source of nutrients such as polyunsaturated fatty acids as well as a major source of ED and other bioaccumulating contaminants, such as methylmercury. Evidence might justify recommendations to increase as well as to reduce fish consumption, quite an uneasy situation for risk managers: decreasing fish consumption (and its nutritional benefits) may not be necessary in Europe, but monitoring of contaminants in edible fish should be continued, as well as the development of novel aquaculture feeds, less liable to contamination [22].

Most important, effects of contaminants and natural food components may interact on the same pathways and targets. The outcomes of interactions may be complex, depending on dose and targets; e.g., phytoestrogens can protect against some hormone-dependent cancers, as well as postmenopausal osteoporosis, but may also interfere with receptor-mediated signal transduction (e.g. by inhibiting protein kinase) and DNA replication [23]. Up to date, scientific data available on interactions between xenobiotics and “natural” substances in food are still limited; below, some relevant examples are provided

Iodine and ED

Iodine is the main determinant of thyroid development and function; seafood and milk are the main dietary sources. Subclinical iodine deficiency is still a common problem in many areas, including Europe [24]; thyroid is also increasingly recognized as a major target for ED, including newly recognized ones, such as organophosphorus insecticides [25]. Yet, only a few papers target low iodine status in relation to susceptibility to xenobiotics. Somewhat unexpectedly phthalates, the widespread plasticizers known mainly as antiandrogens, can modulate basal iodide uptake mediated by the sodium/iodide symporter in thyroid follicular cells *in vitro*: the effect was not shared by all phthalates and was independent from cytotoxicity [26]. Many phytoestrogens may interfere with iodination of thyroid hormones. Some (e.g., naringenin, and quercetin, which contain a resorcinol moiety) are direct and potent inhibitors of thyroid peroxidase, others (myricetin, naringin) show noncompetitive inhibition of tyrosine iodination with respect to iodine ion, whereas biochanin A may act as an alternate substrate for iodination [27]. A Czech biomonitoring study in children also indicated an adverse effect of genistein on thyroid function [28]. The drinking-water contaminant perchlorate inhibits thyroidal iodide uptake; however, iodine-deficient female rats were more resistant to the inhibition of iodine absorption from perchlorate exposure than normal rats [29]. Thus, the

interaction between iodine and some thyroid-targeting ED may be less straightforward than expected.

Phytoestrogens and the “xeno”ED

Due to their pleomorphic biological effects, phytoestrogens are a sort of “natural ED”, whose overall dietary intake of phytoestrogens may be significant also in Europe [23, 30, 31]. Flavonoids (daidzein, genistein, quercetin, and luteolin) can at least partly antagonize the proliferation-stimulating activity of synthetic estrogenic ED in estrogen-dependent MCF-7 human breast cancer cells: these ED included anionic detergent by-products alkylphenols, plastic additive bisphenol A, and the PCB 4-dihydroxybiphenyl [32, 33]. These findings suggest that phytoestrogens can compete with estrogenic ED on shared biological targets, thus exerting a protective action. In other models no interaction was observed: genistein did not modulate the effects on human astroglial cells by two persistent ED, the polybrominated flame retardant PBDE-99 and the PCB mixture Aroclor 1254 [34]. As it is sometimes the case, *in vivo* studies provide a more complex picture. Genistein and the estrogenic chlorinated insecticide methoxychlor had an additive impact on both immune function and immune functional development in rats; the developing thymus appeared as a sensitive target of combined exposure [35]. In estrogen reporter (ERE-tK-Luciferase) male mice genistein modulated the actions of both estradiol and persistent ED in liver and testis with tissue-specific features: the antiestrogenic action of beta-hexachlorocyclohexane in the testis and o,p'-DDT in the liver was antagonized, whereas genistein had an additive effect with the ER agonist p,p'-DDT in the liver [36]. Two predefined mixtures of phytoestrogens and synthetic ED were tested in the uterotrophic assay on prepubertal rats: the composition of each mixture (what chemicals and to what amount) was based on human exposure data. The phytoestrogen mixture did elicit an uterotrophic response, whereas the synthetic one has no effect itself nor an additive effect with phytoestrogens, possibly because of exposure levels too low [37]. The combined exposure to estrogenic and antiandrogenic ED is suggested as a potential risk to male reproductive development. Genistein and the antiandrogenic fungicide vinclozolin, alone or in combination, were investigated concerning the induction of hypospadias in mice: the incidences were 25%, 42% and 41% for genistein, vinclozolin and combined treatment, respectively, indicating a less than additive effect [38]. On the other hand, genistein, as well as the methyl donor folic acid, both antagonized the DNA hypomethylating effect of bisphenol A in mouse embryos [39]. The available data indicate that interactions between phytoestrogens and ED can be important, but cannot simply be explained in terms of additivity or antagonism; indeed, additivity and antagonism may vary, depending on the chemicals, endpoints and lifestages.

ED and Vitamin A Pathways

Retinoic acid is the internal form of vitamin A interacting with the nuclear receptors RAR and RXR, whose natural ligands are all-trans-retinoic acid and 9-cis-retinoic acid, respectively. Retinoic acid pathways cross-talk with those of the aryl hydrocarbon receptor (AhR), the direct cell target for dioxins and dioxin-like compounds [40]. Dioxins are potent

inducers of cytochrome P450 (CYP) 1A1, that in its turn may enhance the dioxin effects; the concurrent supplementation of vitamin A inhibits CYP1A1 activity in dioxin-exposed mice, reducing liver damage as well as CYP1A1 and AhR mRNA expression [41]. Mice lacking retinoid binding proteins were especially responsive to dioxin-induced liver retinoid depletion, intracellular retinoid binding protein I being the main factor. RAR- and RXR-knockout mice were essentially sensitive as wild-type mice, with the exception of RXRbeta^{-/-} mice which showed no decrease in hepatic Vitamin A concentration; this suggests a possible role of RXRbeta in dioxin-induced retinoid disruption [42]. Retinoid storage and metabolism were also disrupted in female rats of two strains with different dioxin sensitivity (Long-Evans and Han/Wistar) [43]. Comparison of dioxin effects on liver retinyl palmitate in AhR^{+/-} and AhR^{-/-} mice support disruption of retinoid homeostasis as a primary AhR-mediated mode of action of dioxin-like chemicals [44]. Retinoid pathways can be a critical target also for polybrominated diphenyl ethers: in rats treated orally with pentaBDE-71, decrease of hepatic apolar retinoids was the most sensitive effect, together with reduced thyroid hormone [45]. These studies might also hint to vitamin A deficiency as a susceptibility factor towards some persistent ED.

Although the portfolio of scientific evidence is still quite limited, several other examples can be retrieved from the Endocrine disrupting chemicals – Diet Interaction Database - EDID, the only dedicated database available on ED-nutrient interactions [19]. One further instance is the general protective action elicited by “antioxidant” vitamins C and E towards the effects of several EDs, including dioxin-like polychlorinated biphenyls (PCB) and phthalates; indeed, several ED-related modes of action seem to eventually lead to increased oxidative stress [46]. Overall, new evidence on interactions between ED and natural food components may disclose new insights on food-related factors modulating vulnerability as well as on nutrient intake as support to risk prevention and/or risk reduction strategies.

EXPOSURE TO EDCS AND IMPACT ON THE FETO-PLACENTAL UNIT

Maternal exposure to EDCs has been demonstrated to be a significant reason for increases in adverse pregnancy and fetal outcomes. The placenta protects and nourishes the fetus by regulating nutrient and xenobiotic homeostasis between the maternal and fetal compartments. As discussed below, xenobiotics that can affect this placental homeostatic control may lead to abnormal fetal development by altering fetal exposure to toxic compounds and/or nutrient homeostasis [65, 66, 69]. An important aspect of this review is to highlight some areas in which EDCs have drawn considerable attention due to the many potential fetotoxic effects, which may be caused upon *in utero* exposure. For example, recent evidence suggests a link between EDC exposure and the fetal origins of neurological impairment that cannot be ignored even though their mechanistic basis is not well understood [47-51]. EDCs are hypothesized to induce functional and/or structural changes in specific neuroendocrine pathway(s), effects being largely dependent upon the phase (gestational time) and level of exposure [52]. In addition, the potential for additive or synergistic effects of low dose com-

binations of EDCs are not well established and require considerable investigation. [53]. Moreover, the role of other factors including diet, exercise and genetics has not been well characterized, adding to the difficulty in delineating the role of EDCs on neurodevelopment. Finally, the pharmacokinetic and pharmacodynamic relationships for EDCs differ and there exists a potential for placental and fetal accumulation not accurately measured in maternal plasma [52, 54]. For example, a recent study revealed that when both newborns and adults are exposed to the same bisphenol A (BPA) levels, newborns retain up to 3 times more than adults [55].

BPA is an EDC due to its ability to interact with estrogen receptor (ER α and β) isoforms, androgen receptors (AR), and possessing a high affinity for the estrogen related receptor α (ERR α) during mammalian brain development [56]. BPA eluted from polycarbonate drinking bottles was demonstrated to exert an estrogenic like neurotoxic effect in developing cerebellar neurons [49]. BPA has also been demonstrated to alter fetal neurodevelopment through thyroid hormone (TH) pathways, as recently in a TH-dependent dendritic Purkinje cell development in a murine cerebellar culture assay [51, 57].

BPA is metabolized into BPA glucuronide, which is hormonally inactive, and excreted *via* the urine with a half-life below 6 hours [58]. Despite the rapid metabolism, the U.S. CDC have found BPA levels in 95% of all human urine samples tested, suggesting broad and continuous BPA exposure [59, 60]. Chronic exposure to low levels of BPA may still cause developmental toxicity due to the significant potential for bioaccumulation in the human placenta and fetus [61]. For example, human BPA levels in the placenta and amniotic fluid were 5 folds higher at weeks 15 to 18 compared to maternal serum [62]. Furthermore, fetal and perinatal exposure to BPA has been linked to several neurodevelopmental changes and disorders including autism and the related autism spectrum of disorders (ASD), schizophrenia, impaired neurotransmission, attention deficit and hyperactivity disorders, and potentially sexual dimorphic related changes in brain structure and function, as recently reviewed by Brown [51].

Phthalates are a ubiquitous class of environmental teratogens capable of exerting their toxic effects through several nuclear hormone receptors including the androgen receptor (AR) antagonism [52, 63], ER agonism [52, 64] and/or *trans*-activation of the peroxisome-proliferator activated receptors (PPAR) α and γ isoforms, either directly or indirectly [65-69]. Phthalates are classified as peroxisome proliferator chemicals due to their effects on peroxisomal lipid metabolism [70]. Moreover, they may also exert their neurotoxic effects through altering zinc metabolism [71, 72] or by altering intracellular Ca²⁺ concentrations leading to the formation of reactive oxygen species potentially through a protein kinase C mediated pathway [73]. It has also been suggested that DEHP inhibits membrane Na⁺-K⁺ ATPase in the rat brain, a phenomena linked to several neurodegenerative and psychiatric disorders [74].

Phthalate reproductive toxicology research [63, 65-69] has been largely focused on di-(2-ethylhexyl)-phthalate

(DEHP), an industrial plasticizer that is ubiquitously dispersed in the environment. Human DEHP exposure most likely begins in the mother's womb, where DEHP has been demonstrated to readily cross the placenta and accumulate in the fetus [67, 68]. DEHP mediated direct or indirect PPAR effects on placental essential fatty acid (EFA) homeostasis have also been of interest [64-69]. The fetus requires maternal dietary intake and placental transfer of EFAs to guide proper pregnancy outcomes and fetal development, e.g. neurodevelopment [75-78]. The placenta plays a fetoprotective role by accumulating EFAs from maternal circulation and directionally transporting them into fetal compartment [79, 80]. With regards to proper neurodevelopment, EFAs including docosahexaenoic acid (DHA, 22:6n-3) and arachidonic acid (AA, 20:4n-6) are known to play critical roles in myelogenesis and serve as essential components in neurogenesis, thus making the fetoprotective role of the placenta essential for neurodevelopment [76-80]. EFA imbalances have been linked to several neurological disorders including autism and ASDs, bipolar disorder, and schizophrenia, suggesting that a proper EFA supply is required to protect the CNS development [81, 82].

PPAR α and γ regulate the expression of several fatty acid transport conferring proteins and metabolizing enzymes that maintain essential fatty acid (EFA) homeostasis and can be *trans*-activated by DEHP and its metabolites, mono-(2-ethylhexyl)-phthalate (MEHP) and 2-ethylhexanoic acid (EHA) [65, 66, 83]. Recent studies revealed that DEHP and its metabolites MEHP and EHA, can significantly increase the expression of EFA homeostasis proteins, EFA and lipid accumulation in the lipid metabolome of HRP-1 *in vitro* rat placental cell line [65, 84]. It was also demonstrated that the resulting increase in the expression of fatty acid transport-conferring proteins in these cells also led to a significant increase in fatty acid and lipid accumulation in the cells [85]. DEHP exposure has been revealed to alter the expression of EFA homeostasis proteins in the *in vivo* rat placenta [66]. In this study, radiolabeled AA and DHA were administered to rat dams at gestational day (GD) 20 and the maternal, placental and fetal disposition of the labeled EFAs were assessed [66]. AA was significantly reduced in the maternal and fetal plasma, yet increased significantly in the placenta upon DEHP exposure. DHA levels significantly increased in the maternal plasma and decreased in the fetal plasma upon DEHP exposure contrasted to the control vehicle. DEHP exposure also elicited a statistically significant decrease of both AA and DHA in the developing fetal brain. Lipomic analysis also revealed that DEHP reduced fetal pup brain accumulation of several critical fatty acid and lipid classes including a significant decrease in sphingomyelin (SM) of 54% [69]. DEHP exposure elicited a significant reduction of DHA in five lipid fractions (namely, cholesterol ester (CE), diacylglyceride, phosphatidylserine, lysophosphatidyl choline (LYPC) and SM), whereas AA was significantly decreased CE and LYPC. SM and DHA levels are critical for proper brain development including neurogenesis and neuronal differentiation [76-78]. Moreover, the brain weight and active neurogenesis rapidly increases from GD15 to term in the rat fetus [79]. These results suggest that DEHP may adversely affect on fetal neurodevelopment.

Recently, it has been found that 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) can reduce n-3 and n-6 EFAs in contrast to the control when administered to the cynomolgus macaque at GD15 or 20 and the brains were isolated at GD24-26 [85]. Although the mechanism of action was not defined, improper neural tube closing and other neurodevelopmental aberrations were observed and attributed to the improper EFA balance [86-89]. Interestingly, TCDD also has an estrogenic response, acting through ERs, potentially interacting with the aryl hydrocarbon receptor (AHR) in rats [90, 91].

In summary, several EDCs have been demonstrated to exert their teratogenic effects on fetal neurodevelopment. Considerable attention is necessary to elucidate the mechanisms by which individual or multiple combinations of EDCs can elicit fetal neurotoxicity. The extent to which the animal data may be extrapolated to predict a human response will also need to be determined.

ED EXPOSURE AND PAEDIATRIC ENDOCRINE DISORDERS

Normal human sex differentiation, growth and puberty are critically dependent upon hormonal actions, opening up for targeting by EDCs. Recent epidemiological studies demonstrate increasing incidences of related developmental disorders in children originating in embryonic and fetal life [92]. Environmental influences may play an important role in the pathogenesis of such disorders although the factors involved remain to be characterized [7,8].

Disorders of Sex Development (DSD)

It has been hypothesized that certain defined disorders of male sex differentiation may be linked by common pathogenic mechanisms [92]. Cryptorchidism, hypospadias, testicular cancer and poor semen quality are all proposed to be components of this "syndrome". It has been suggested that exposure to environmental estrogens or anti-androgens may alter the fetal hormonal balance, or compete directly with the androgen receptor, causing undermasculinization of male fetuses. A severe outcome of such action would potentially result in DSD with an intersex phenotype at birth. Less severe disruption may cause maldescended testes or hypospadias. More recently, epigenetic alterations with transgenerational influences have been implicated in the effects of EDCs on reproductive functions [93].

Cryptorchidism

Cryptorchidism is difficult to study due to poor reporting but certain investigations point to an increased incidence in some countries during several decades [94]. Model substances, particularly antiandrogens, have been associated with cryptorchidism in experimental animals [94]. Prenatal exposure to pesticides (as indicated from concentrations in breast milk) has been linked with an increased risk of cryptorchidism [95]. Higher organochlorine concentrations were found in fat samples from cryptorchid boys when compared with control samples [96] but other studies have failed to demonstrate similar correlations [97]. Although data is accumulating, at the present time no single EDC can be convincingly blamed for causing cryptorchidism.

Hypospadias

Hypospadias is underreported to malformation registries, mainly since the most common mild cases are not reported. Still there are reports from reliable sources indicating an increased incidence in European countries. Large regional variation seems to exist [98, 99]. Similarly to cryptorchidism, exposure studies investigating links to EDCs have come to opposing results and it is fair to say that at the present stage of knowledge the data are inconclusive, mainly due to low power and poor disease classification of most published studies.

Puberty

The onset and tempo of puberty are under endocrine control and there is evidence to indicate that EDCs may affect pubertal development [100]. The proposed adverse effects of EDCs are most often related to premature thelarche in girls. Studies in the US have demonstrated a recent trend to an earlier start of puberty in girls, particularly in certain ethnic groups, with a direct correlation to obesity, as assessed as Body Mass Index (BMI). Exposure to EDCs has been proposed to play a role in this novel trend [101]. One group recently found a similar development in Denmark but failed to show a correlation with BMI and levels of reproductive hormones [102]. This indicates an influence by environmental factors, the natures of which are yet to be determined.

Thyroid

EDCs may exert unwanted actions on thyroid function [103]. Links between exposure to PCBs and increased thyroid stimulating hormone have been observed by some [104, 105], but not other authors [106-107]. PCB has been proposed to exert goiterogenic actions [108], and pentachlorophenol indicated to suppress thyroid hormone levels in newborns [109]. Given the critical role of the thyroid for fetal and infant neurodevelopment, such action may be associated with adverse neurodevelopmental consequences in children.

Adrenal

EDCs are known to exert harmful actions in the adrenal cortex. DDT metabolites are well known inhibitors of adrenal function [110], exerting direct cytotoxicity to adrenocortical cells. Certain EDCs may have a negative impact on adrenal steroidogenesis. The phytoestrogen resveratrol was found to suppress glucocorticoid production by inhibiting 21-hydroxylase [5]. This may impair glucocorticoid driven stress responses, which is a crucial component of the host-defense system against infection and injury.

ED AS POSSIBLE RISK FACTORS IN ENDOCRINE AUTOIMMUNITY

Autoimmune endocrine disorders are diseases of the endocrine glands caused by an impaired response of the immune system, which fails to recognize self-antigens and reacts against them [111]. Most autoimmune endocrine disorders target single organs, but often patients have multiple autoimmune endocrine disorders or autoimmune polyglandular syndromes, where, two or more endocrine glands are involved by the autoimmune process [112]. Autoimmune endocrine disorders include: type 1 diabetes mellitus, Hashi-

moto's thyroiditis, Graves' disease, Addison's disease, autoimmune hypophysitis, autoimmune oophoritis, autoimmune hypoparathyroidism, testicular insufficiency, and premature ovarian failure.

Autoimmune diseases are usually the result of an interplay between genetic and acquired factors; the latter including also environmental influences [112]. Autoimmune endocrine diseases often present a sexual dimorphism, which suggests a potential role of sexual hormones on the immune system and therefore their contribution to autoimmune disorders [113].

Several studies have reported that sex steroid hormones can modulate the immune system [114]. Although, it is important to bear in mind that the response to sex hormones can vary among individuals, in relation to age, genetic background, duration and level of exposure; in general estrogens and androgens appear to have opposite effects on the immune system [115]. In particular, estrogen treatment in mice has been associated with reduced number of lymphocytes in several organs as well as with a deregulation in the balance between T and B lymphocytes [115]. The most common finding has been an estrogen-related induction of B-cell hyperactivity and T-cell hypoactivity [115]. This enhanced immunoreactivity in females has been defined as a 'double-edged sword', which on the one hand protects against infections, whereas on the other hand increases the risk of developing autoimmune diseases [116].

There is growing evidence suggesting that EDCs can affect the immune system, promoting the development of autoimmune diseases [117]. An example is given by environmental estrogens, which could exert the same effect than endogenous estrogens on the immune system [113]. EDCs could act on the immune system through different mechanisms [118]: 1) inhibiting the processes involved in establishing tolerance, with generation of autoreactive cells; 2) modifying gene expression in cells involved in the immune response, permitting lymphocytes to respond to signals normally insufficient to initiate a response or permitting the antigen-presenting cells to abnormally stimulate a response; 3) modifying self-molecules such that they are recognized by the immune system as foreign.

Yurino *et al.* [119] have demonstrated that EDCs, such as diethylstilbestrol and bisphenol-A (BPA), can stimulate autoantibody production by B1 cells both *in vitro* and *in vivo*. The majority of data related to ED and autoimmunity are on animal models of lupus [114]. It has been shown that prolactin or estrogen levels accelerate disease activity in lupus-prone mice or can induce a lupus-like syndrome in normal mice. Scant data are available on the effect of ED specifically on endocrine autoimmune diseases [120] and therefore, this area needs further explorations.

BIOLOGICAL MONITORING OF ENDOCRINE DISRUPTORS AND ASSESSMENT OF HEALTH RISKS: THE CASE OF PHTHALATES

Over the years the need for risk assessments has increased because of the potential health risks related to exposure to EDs in everyday life worldwide. Biological monitoring, defined as measuring concentrations of chemicals and

their metabolites in human body fluids, has been shown to represent an accurate, efficient and cost-effective exposure assessment to environmental pollutants even at low levels [121]. By detecting non-invasive biomarkers through appropriate analytical procedures, human bio-monitoring (HBM) takes into account all routes and sources of exposure, thus representing an ideal instrument for risk assessment and risk management [122, 123]. The sensitivity of HBM methods enables the elucidation of the metabolism and modes of action of pollutants in humans. The diesters of benzene-1,2-dicarboxylic (phthalic) acid, commonly known as phthalates, are a family of industrial compounds with a common chemical structure, dialkyl or alkyl/aryl esters of 1,2-benzenedicarboxylic acid [124]. They are high-production-volume synthetic chemicals and the probability of exposure to these chemicals is high because of their use in plastic items and other common consumer products, including personal-care products (e.g., perfumes, lotions, cosmetics), paint, industrial plastics, medical devices and pharmaceuticals; phthalates are primarily used as plasticizers to impart flexibility to an otherwise rigid polyvinylchloride (PVC) [124]. As these plasticizers are not chemically bound to PVC, they elute at a constant rate from plastic products into the environment. Phthalates have been found everywhere, even in infant formulas and breast milk [125, 126].

Recently, food has been observed to be the predominant intake source of DEHP, whilst other sources, such as enteric coatings in medications, considerably contributed to the daily intake of di-n-butyl phthalate (DnBP) and diisobutyl phthalate (DiBP) in an adult population [127, 128]. A comparison of the available data resulting from the determination of the target compound in indoor air and house dust as well as emission studies with the results from the HBM studies reveals that only a small portion of intake takes place *via* the air, dust paths and personal care products, such as cosmetics [129, 130].

Phthalates are rapidly metabolized to their monoesters, which can be further transformed into oxidative metabolites, conjugated, and both free and conjugated metabolites excreted in the urine and faeces [131].

Due to their chemical properties exposure to phthalates does not result in bioaccumulation [132].

Urinary secondary metabolites have been shown to represent ideal biomarkers of exposure to phthalates, allowing accurate assessments of human exposure from multiple sources and routes; the use of metabolites also avoids the analytical problems caused by the risk of sample's contamination by the ubiquitous parent phthalates [133-135].

As there may be significant demographic variations in exposure and/or metabolism of phthalates, health-risk assessments for phthalate exposure in humans should consider different potential risk groups [136].

During the last decades, a great deal of scientific and public concern has been raised about the potential health hazards posed by exposures to phthalates, even in environmental concentrations [137-144].

In particular, phthalates with side-chain lengths C4 to C6 are known to adversely affect the differentiation and function of the reproductive system [145-149].

The Center for the Evaluation of Risks to Human Reproduction (CERHR) identified two specific situations as potentially problematic, the exposure of young children to diisononyl phthalate (DINP) through the use of toys or to DEHP from medical devices [150].

To this regard, exposure assessment have observed that the exposure of children to phthalates exceeds that in adults and the tolerable intake of children is frequently exceeded, in some instances up to 20-fold [151,152].

On the other hand, newborns in the Neonatal Intensive Care Unit (NICU) environment represent a population at particularly increased risk for exposure to DEHP, because of their physical conditions, small body size and contemporary exposure to multiple medical devices containing DEHP. To this regard, it has been documented that these newborns can be exposed up to 100 times above the limit values, depending on the intensity of medical care [153, 154]. As a consequence, the U.S.A and European Union promulgated limitations of use for certain phthalates [155]. Although several risk assessments have been finalized for these chemicals during the last decade, in the future there is the need for risk assessments able to cover a high number of exposure situations and a transparent process of collecting data, thus ensuring the safety of workers and consumers [156].

CONCLUSIONS

There is accumulating evidence suggesting that EDCs represent a large and heterogeneous group of chemical compounds that may potentially affect human health, especially if exposures occur at early stages in life, including both prenatal life and early childhood. Effects are especially relevant in relation to the long-term development of the reproductive and nervous systems, as well as for metabolic programming.

Factors co-modulating the risk, besides age and gender, include diet and lifestyle.

As a consequence, in the future risk assessment of human health should include: (i) population-based estimates of environmental total exposure levels from several sources using HBM; (ii) further studies to assess interactions between several different classes of ubiquitous compounds and their combined effect; (iii) characterization of appropriate biomarkers of effective dose and susceptibility for major groups of EDCs.

ACKNOWLEDGEMENTS

Part of the work has been performed within the frame of the activities of the Network of Excellence CASCADE (6th framework programme, www.cascadenet.org <<http://www.cascadenet.org>>).

The authors gratefully acknowledge the support of Mrs. Francesca Baldi (Istituto Superiore di Sanità - Roma, Italy) in the preparation of the manuscript.

REFERENCES

- [1] Caserta, D.; Maranghi, L.; Mantovani, A.; Marci, R.; Maranghi, F.; Moscarini, M. Impact of endocrine disruptor chemicals in gynaecology. *Hum. Reprod. Updat.* **2008**, *14*, 59-72.

- [2] Landrigan, P.; Garg, A.; Droller, D.B. Assessing the effects of endocrine disruptors in the National Children's Study. *Environ. Health Perspect.*, **2003**, *111*, 1678-1682.
- [3] Svechnikov, K.; Svechnikova, I.; Söder, O. Inhibitory effects of mono-ethylhexyl phthalate on steroidogenesis in immature and adult rat Leydig cells *in vitro*. *Reprod. Toxicol.*, **2008**, *25*, 485-490.
- [4] Svechnikova, I.; Svechnikov, K.; Söder, O. The influence of di-(2-ethylhexyl) phthalate on steroidogenesis by the ovarian granulosa cells of immature female rats. *J. Endocrinol.*, **2007**, *194*, 603-609.
- [5] Supornsilchai, V.; Svechnikov, K.; Seidlova-Wuttke, D.; Wuttke, W.; Söder, O. Phytoestrogen resveratrol suppresses steroidogenesis by rat adrenocortical cells by inhibiting cytochrome P450 c21-hydroxylase. *Horm. Res.*, **2005**, *64*, 280-286.
- [6] Latini, G.; Scoditti, E.; Verrotti, A.; De Felice, C.; Massaro, M. Peroxisome proliferator-activated receptors as mediators of phthalate-induced effects in the male and female reproductive tract: epidemiological and experimental evidence. *PPAR. Res.*, **2008**, *2008*, 359-367.
- [7] Pombó, M.; Castro-Feijóo, L. Endocrine disruptors. *J. Pediatr. Endocrinol. Metab.*, **2005**, *18*(Suppl 1), 1145-1155.
- [8] Waring, R.H.; Harris, R.M. Endocrine disruptors: a human risk? *Mol. Cell. Endocrinol.*, **2005**, *244*, 2-9.
- [9] Whitehead, S.A.; Rice, S. Endocrine-disrupting chemicals as modulators of sex steroid synthesis. *Best Pract. Res. Clin. Endocrinol. Metab.*, **2006**, *20*, 45-61.
- [10] Henley, D.V.; Korach, K.S. Endocrine-disrupting chemicals use distinct mechanisms of action to modulate endocrine system function. *Endocrinology.*, **2006**, *147*(Suppl 6), 25-32.
- [11] Iguchi, T.; Watanabe, H.; Katsu, Y. Application of ecotoxicogenomics for studying endocrine disruption in vertebrates and invertebrates. *Environ. Health Perspect.*, **2006**, *114*(Suppl 1), 101-105.
- [12] Schulte-Oehlmann, U.; Albanis, T.; Allera, A.; Bachmann, J.; Berntsson, P.; Beresford, N.; Carnevali, D.C.; Ciceri, F.; Dagnac, T.; Falandysz, J.; Galassi, S.; Hala, D.; Janer, G.; Jeannot, R.; Jobling, S.; King, I.; Klingmüller, D.; Kloas, W.; Kusk, K.O.; Levada, R.; Lo, S.; Lutz, I.; Oehlmann, J.; Oredsson, S.; Porte, C.; Rand-Weaver, M.; Sakkas, V.; Sugni, M.; Tyler, C.; van Aarle, R.; van Ballegoy, C.; Wollenberger, L. Compendo: Focus and approach. *Environ. Health Perspect.*, **2006**, *114*(Suppl 1), 98-100.
- [13] Gore, A.C. Developmental programming and endocrine disruptor effects on reproductive neuroendocrine systems. *Front. Neuroendocrinol.*, **2008**, *29*, 358-374.
- [14] Crews, D.; Gore, A.C.; Hsu, T.S.; Dangleben, N.L.; Spinetta, M.; Schallert, T.; Anway, M.D.; Skinner, M.K. Transgenerational epigenetic imprints on mate preference *Proc. Natl. Acad. Sci. U S A.*, **2007**, *104*, 5942-5946.
- [15] Hokanson, R.; Hanneman, W.; Hennessey, M.; Donnelly, K.C.; McDonald, T.; Chowdhary, R.; Busbee, D.L. DEHP, bis(2-ethylhexyl phthalate, alters gene expression in human cells: possible correlation with initiation of fetal developmental abnormalities. *Hum. Exp. Toxicol.*, **2006**, *25*, 687-695.
- [16] Willingham, E. Endocrine-disrupting compounds and mixtures: unexpected dose-response. *Arch. Environ. Contam. Toxicol.*, **2004**, *46*, 265-269.
- [17] Fromme, H.; Albrecht, M.; Angerer, J.; Drexler, H.; Gruber, L.; Schlummer, M.; Parlar, H.; Korner, W.; Wanner, A.; Heitmann, D.; Roscher, E.; Bolte, G. Integrated Exposure Assessment Survey (INES) exposure to persistent and bioaccumulative chemicals in Bavaria, Germany. *Int. J. Hyg. Environ. Health.*, **2007**, *210*, 345-349.
- [18] European Food Safety Authority. Opinion of the Scientific Panel on Contaminants in the Food Chain on a request from the Commission to assess the health risks to consumers associated with exposure to organotins in foodstuffs. *EFSA J.*, **2004**, *102*, 1-119.
- [19] Baldi, F.; Mantovani, A. A new database for food safety: EDID (Endocrine disrupting chemicals - Diet Interaction Database). *Ann. Ist. Super. Sanità.*, **2008**, *44*, 57-63.
- [20] Fattore, E.; Fanelli, R.; Turrini, A.; Di Domenico, A. Current dietary exposure to polychlorodibenzo-p-dioxins; polychlorodibenzofurans and dioxin-like polychlorobiphenyls in Italy. *Mol. Nutr. Food Res.*, **2006**, *50*, 915-921.
- [21] Mozaffarian, D.; Rimm, E.B. Fish intake, contaminants and human health: evaluating the risks and the benefits. *JAMA*, **2006**, *296*, 1885-1899.
- [22] European Food Safety Authority. Opinion of the Scientific Panel on Contaminants in the food chain on a request from the European Parliament related to the safety assessment of wild and farmed fish. *EFSA J.*, **2005**, *236*, 1-118.
- [23] Martin, J.H.; Crotty, S.; Nelson, P.N. Phytoestrogens: perpetrators or protectors? *Future Oncol.*, **2007**, *3*, 307-318.
- [24] European Food Safety Authority. Opinion of the Scientific Panel on Additives and Products or Substances used in Animal Feed on the request from the Commission on the use of iodine in feedstuffs. *EFSA J.*, **2005**, *168*, 1-42.
- [25] De Angelis, S.; Tassinari, R.; Maranghi, F.; Eusepi, A.; Di Virgilio, A.; Chiarotti, F.; Ricceri, L.; Venerosi Pesciolini, A.; Gilardi, E.; Moracci, G.; Calamandrei, G.; Olivieri, A.; Mantovani, A. Developmental exposure to chlorpyrifos induces alterations in thyroid and thyroid hormone levels without other toxicity signs in cd1 mice. *Toxicol. Sci.*, **2009**, *108*, 311-319.
- [26] Wenzel, A.; Franz, C.; Breous, E.; Loos, U. Modulation of iodide uptake by dialkyl phthalate plasticizers in FRTL-5 rat thyroid follicular cells. *Mol. Cell. Endocrinol.*, **2005**, *244*, 63-71.
- [27] Divi, R.L.; Doerge, D.R. Inhibition of thyroid peroxidase by dietary flavonoids. *Chem. Res. Toxicol.*, **1996**, *9*, 16-23.
- [28] Milerová, J.; Cerovská, J.; Zamrazil, V.; Břelk, R.; Lapčík, O.; Hampl, R. Actual levels of soy phytoestrogens in children correlate with thyroid laboratory parameters. *Clin. Chem. Lab. Med.*, **2006**, *44*, 171-174.
- [29] Paulus, B.F.; Bazar, M.A.; Salice, C.J.; Mattie, D.R.; Major, M.A. Perchlorate inhibition of iodide uptake in normal and iodine-deficient rats. *J. Toxicol. Environ. Health A.*, **2007**, *70*, 1142-1149.
- [30] Saarinen, N.M.; Bingham, C.; Lorenzetti, S.; Mortensen, A.A.; Mäkela, S.; Penttinen, P.; Sorensen, I.K.; Valsta, L.M.; Virgili, F.; Vollmer, G.; Wärrä, A.; Zierau, O. Tools to evaluate estrogenic potency of dietary phytoestrogens: a consensus paper from the EU thematic network "Phytohealth" (QLKI-2002-2453). *Genes Nutr.*, **2006**, *1*, 143-158.
- [31] Peeters, P.H.; Slimani, N.; van der Schouw, Y.T.; Grace, P.B.; Navarro, C.; Tjonneland, A.; Olsen, A.; Clavel-Chapelon, F.; Touillaud, M.; Boutron-Ruault, M.C.; Jenab, M.; Kaaks, R.; Linseisen, J.; Trichopoulos, A.; Trichopoulos, D.; Dilis, V.; Boeing, H.; Weikert, C.; Overvad, K.; Pala, V.; Palli, D.; Panico, S.; Tumino, R.; Vineis, P.; Bueno-de-Mesquita, H.B.; van Gils, C.H.; Skeie, G.; Jakszyn, P.; Hallmans, G.; Berglund, G.; Key, T.J.; Travis, R.; Riboli, E.; Bingham, S.A. Variations in plasma phytoestrogen concentrations in European adults. *J. Nutr.*, **2007**, *137*, 1294-300.
- [32] Han, D.H.; Denison, M.S.; Tachibana, H.; Yamada, K. Relationship between estrogen receptor-binding and estrogenic activities of environmental estrogens and suppression by flavonoids. *Biosci. Biotechnol. Biochem.*, **2002**, *66*, 1479-1487.
- [33] Rajapakse, N.; Silva, E.; Scholze, M.; Kortenkamp, A. Deviation from additivity with estrogenic mixtures containing 4-nonylphenol and 4-tert-octylphenol detected in the E-SCREEN assay. *Environ. Sci. Technol.*, **2004**, *38*, 6343-6352.
- [34] Madia, F.; Giordano, G.; Fattori, V.; Vitalone, A.; Branchi, I.; Capone, F.; Costa, L.G. Differential *in vitro* neurotoxicity of the flame retardant PBDE-99 and of the PCB Aroclor 1254 in human astrocytoma cells. *Toxicol. Lett.*, **2004**, *154*, 11-21.
- [35] Guo, T.L.; Zhang, X.L.; Bartolucci, E.; McCay, J.A.; White, K.L. Jr.; You, L. Genistein and methoxychlor modulate the activity of natural killer cells and the expression of phenotypic markers by thymocytes and splenocytes in F0 and F1 generations of Sprague-Dawley rats. *Toxicology*, **2002**, *172*, 205-215.
- [36] Penza, M.; Montani, C.; Romani, A.; Vignolini, P.; Ciana, P.; Maggi, A.; Pampaloni, B.; Caimi, L.; Di Lorenzo, D. Genistein accumulates in body depots and is mobilized during fasting: reaching estrogenic levels in serum that counter the hormonal actions of estradiol and organochlorines. *Toxicol. Sci.*, **2007**, *97*, 299-307.
- [37] van Meeuwen, J.A.; van den Berg, M.; Sanderson, J.T.; Verhoef, A.; Piersma, A.H. Estrogenic effects of mixtures of phyto- and synthetic chemicals on uterine growth of prepubertal rats. *Toxicol. Lett.*, **2007**, *170*, 165-176.
- [38] Vilela, M.L.; Willingham, E.; Buckley, J.; Liu, B.C.; Agras, K.; Shiroyanagi, Y.; Baskin, L.S. Endocrine disruptors and hypospadias: role of genistein and the fungicide vinclozolin. *Urology*, **2007**, *70*, 618-621.

- [39] Dolinoy, D.C.; Huang, D.; Jirtle, R.L. Maternal nutrient supplementation counteracts bisphenol A-induced DNA hypomethylation in early development. *Proc. Natl. Acad. Sci. USA*, **2007**, *104*, 13056-13061.
- [40] Murphy, K.A.; Quadro, L.; White, L.A. The intersection between the aryl hydrocarbon receptor (AhR) and retinoic acid-signaling pathways. *Vitam. Horm.*, **2007**, *75*, 33-67.
- [41] Yang, Y.M.; Huang, D.Y.; Liu, G.F.; Zhong, J.C.; Du, K.; Li, Y.F.; Song, X.H. Inhibitory effects of vitamin A on TCDD-induced cytochrome P-450 1A1 enzyme activity and expression. *Toxicol. Sci.*, **2005**, *85*, 727-734.
- [42] Hoegberg, P.; Schmidt, C.K.; Fletcher, N.; Nilsson, C.B.; Trossvik, C.; Gerlienke Schuur, A.; Brouwer, A.; Nau, H.; Ghyselincx, N.B.; Chambon, P.; Hakansson, H. Retinoid status and responsiveness to 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) in mice lacking retinoid binding protein or retinoid receptor forms. *Chem. Biol. Interact.*, **2005**, *156*, 25-39.
- [43] Fletcher, N.; Giese, N.; Schmidt, C.; Stern, N.; Lind, P.M.; Viluksela, M.; Tuomisto, J.T.; Tuomisto, J.; Nau, H.; Hakansson, H. Altered retinoid metabolism in female Long-Evans and Han/Wistar rats following long-term 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD)-treatment. *Toxicol. Sci.*, **2005**, *86*, 264-272.
- [44] Nishimura, N.; Yonemoto, J.; Miyabara, Y.; Fujii-Kuriyama, Y.; Tohyama, C. Altered thyroxine and retinoid metabolic response to 2,3,7,8-tetrachlorodibenzo-p-dioxin in aryl hydrocarbon receptor-null mice. *Arch. Toxicol.*, **2005**, *79*, 260-267.
- [45] van der Ven, L.T.; van de Kuil, T.; Verhoef, A.; Leonards, P.E.; Slob, W.; Cantón, R.F.; Germer, S.; Hamers, T.; Visser, T.J.; Litens, S.; Hakansson, H.; Fery, Y.; Schrenk, D.; van den Berg, M.; Piersma, A.H.; Vos, J.G. A 28-day oral dose toxicity study enhanced to detect endocrine effects of a purified technical pentabromodiphenyl ether (pentaBDE) mixture in Wistar rats. *Toxicology*, **2008**, *245*, 109-122.
- [46] Zhou, C.; Zhang, C. Protective effects of antioxidant vitamins on Aroclor 1254-induced toxicity in cultured chicken embryo hepatocytes. *Toxicol. In vitro*, **2005**, *19*, 665-673.
- [47] Birnbaum, L. S. Developmental effects of dioxins and related endocrine disrupting chemicals. *Toxicol. Lett.*, **1995**, *82-83*, 743-750.
- [48] Palanza, P.; Gioiosa, L.; vom Saal, F. S.; Parmigiani, S. Effects of developmental exposure to bisphenol A on brain and behavior in mice. *Environ. Res.*, **2008**, *108*, 150-157.
- [49] Le, H. H.; Carlson, E. M.; Chua, J. P.; Belcher, S. M. Bisphenol A is released from polycarbonate drinking bottles and mimics the neurotoxic actions of estrogen in developing cerebellar neurons. *Toxicol. Lett.*, **2008**, *176*, 149-156.
- [50] Nakamura, K.; Itoh, K.; Sugimoto, T.; Fushiki, S. Prenatal exposure to bisphenol A affects adult murine neocortical structure. *Neurosci. Lett.*, **2007**, *420*, 100-105.
- [51] Brown Jr., J. S. Effects of Bisphenol-A and Other Endocrine Disruptors Compared With Abnormalities of Schizophrenia: An Endocrine-Disruption Theory of Schizophrenia. *Schizophr. Bull.*, **2009**, *35*, 256-278.
- [52] Yang, M.; Park, M. S.; Lee, H. S. Endocrine disrupting chemicals: human exposure and health risks. *J. Environ. Sci. Health, Part C.*, **2006**, *24*, 183-224.
- [53] Kortenkamp, A. Ten years of mixing cocktails: a review of combination effects of endocrine-disrupting chemicals. *Environ. Health Perspect.*, **2007**, *115*(Suppl 1), 98-105.
- [54] Coles, L. S.; Eddington, N. D. *AAPS Newsmagazine*, **2007**, *10*, 18.
- [55] Hans, M.; Ursula, G.-R. Bisphenol A levels in blood depend on age and exposure. *Toxicol. Lett.*, **2009**, [Epub June 26, 2009].
- [56] Takayanagi, S.; Tokunaga, T.; Liu, X.; Okada, H.; Matsushima, A.; Shimohigashi, Y. Endocrine disruptor bisphenol A strongly binds to human estrogen-related receptor gamma (ERRgamma) with high constitutive activity. *Toxicol. Lett.*, **2006**, *167*, 95-105.
- [57] Kimura-Kuroda, J.; Nagata, I.; Kuroda, Y. Disrupting effects of hydroxy-polychlorinated biphenyl (PCB) congeners on neuronal development of cerebellar Purkinje cells: a possible causal factor for developmental brain disorders? *Chemosphere*, **2007**, *67*, S412-420.
- [58] Völkel, W.; Colnot, T.; Csanády, G. A.; Filser, J. G.; Dekant, W. Metabolism and kinetics of bisphenol A in humans at low doses following oral administration. *Chem. Res. Toxicol.*, **2002**, *15*, 1281-1287.
- [59] Calafat, A. M.; Kuklenyik, Z.; Reidy, J. A.; Caudill, S. P.; Ekong, J.; Needham, L. L. Urinary concentrations of bisphenol A and 4-nonylphenol in a human reference population. *Environ. Health Perspect.*, **2005**, *113*, 391-395.
- [60] Stahlhut, R. W.; Welshons, W. V.; Swan, S. H. Bisphenol A data in NHANES suggest longer than expected half-life, substantial non-food exposure, or both. *Environ. Health Perspect.*, **2009**, *117*, 784-789.
- [61] Schönfelder, G.; Wittfoht, W.; Hopp, H.; Talsness, C. E.; Paul, M.; Chahoud, I. Parent bisphenol A accumulation in the human maternal-fetal-placental unit. *Environ. Health Perspect.*, **2002**, *110*, A703-707.
- [62] Zoeller, R. T.; Bansal, R.; Parris, C. Bisphenol-A, an environmental contaminant that acts as a thyroid hormone receptor antagonist *in vitro*, increases serum thyroxine, and alters RC3/neurogranin expression in the developing rat brain. *Endocrinology*, **2005**, *146*, 607-612.
- [63] Kavlock, R.; Boekelheide, K.; Chapin, R.; Cunningham, M.; Faustman, E.; Foster, P.; Golub, M.; Henderson, R.; Hinberg, I.; Little, R.; Seed, J.; Shea, K.; Tabacova, S.; Ty, R.; Williams, P.; Zacharewski, T. NTP Center for the Evaluation of Risks to Human Reproduction: phthalates expert panel report on the reproductive and developmental toxicity of di(2-ethylhexyl) phthalate. *Reprod. Toxicol.*, **2002**, *16*, 529-653.
- [64] Lu, K. Y.; Tseng, F. W.; Wu, C. J.; Liu, P. S. Suppression by phthalates of the calcium signaling of human nicotinic acetylcholine receptors in human neuroblastoma SH-SY5Y cells. *Toxicology*, **2004**, *200*, 113.
- [65] Xu, Y.; Cook, T. J.; Knipp, G. T. Effects of di-(2-ethylhexyl)-phthalate (DEHP) and its metabolites on fatty acid homeostasis regulating proteins in rat placental HRP-1 trophoblast cells. *Toxicol. Sci.*, **2005**, *84*, 287-300.
- [66] Xu, Y.; Agarwal, S.; Cook, T. J.; Knipp, G. T. Maternal di-(2-ethylhexyl)-phthalate exposure influences essential fatty acid homeostasis in rat placenta. *Placenta*, **2008**, *29*, 962-969.
- [67] Latini, G.; Del Vecchio, A.; Massaro, M.; Verotti, A.; De Felice, C. In utero exposure to phthalates and fetal development. *Curr. Med. Chem.*, **2006**, *13*, 2527-2534.
- [68] Latini, G.; De Felice, C.; Presta, G.; Del Vecchio, A.; Paris, I.; Ruggieri, F.; Mazzeo, P. Status epilepticus and neurodevelopmental outcome at 2 years of age in an extremely low birth weight infant. *Biol. Neonate*, **2003**, *83*, 22-24.
- [69] Xu, Y.; Agarwal, S.; Cook, T. J.; Knipp, G. T. Di-(2-ethylhexyl)-phthalate affects lipid profiling in fetal rat brain upon maternal exposure. *Arch. Toxicol.*, **2007**, *81*, 57-62.
- [70] Cimini, A.; Sulli, A.; Stefanini, S.; Serafini, B.; Moreno, S.; Rossi, L.; Giorgi, M.; Ceru, M. P. Effects of Di-(2-ethylhexyl)phthalate on peroxisomes of liver, kidney and brain of lactating rats and their pups. *Cell Mol. Biol.*, **1994**, *40*, 1063-1076.
- [71] Peters, J. M.; Taubeneck, M. W.; Keen, C. L.; Gonzalez, F. J. Di(2-ethylhexyl) phthalate induces a functional zinc deficiency during pregnancy and teratogenesis that is independent of peroxisome proliferator-activated receptor-alpha. *Teratology*, **1997**, *56*, 311-316.
- [72] Johnson, S. Micronutrient accumulation and depletion in schizophrenia, epilepsy, autism and Parkinson's disease? *Med. Hypotheses*, **2001**, *56*, 641-645.
- [73] Palleschi, S.; Rossi, B.; Diana, L.; Silvestroni, L. Di(2-ethylhexyl)phthalate stimulates Ca(2+) entry, chemotaxis and ROS production in human granulocytes. *Toxicol. Lett.*, **2009**, *187*, 52-57.
- [74] Dhanya, C. R.; Indu, A. R.; Deepadevi, K. V.; Kurup, P. A. Inhibition of membrane Na(+)-K+ ATPase of the brain, liver and RBC in rats administered di(2-ethyl hexyl) phthalate (DEHP) a plasticizer used in polyvinyl chloride (PVC) blood storage bags. *Indian J. Exp. Biol.*, **2003**, *41*, 814-820.
- [75] Latini, G. Potential hazards of exposure to di-(2-ethylhexyl)-phthalate in babies: a review. *Biol. Neonate*, **2000**, *78*, 269-276.
- [76] Jumpson, J.; Clandinin, M. T. *Brain Development: Relationship to Dietary Lipid and Lipid Metabolism*. AOCS Press, Champaign, IL, **1995**.
- [77] Wainright, P. E. Dietary essential fatty acids and brain function: a developmental perspective on mechanisms. *Proc. Nutr. Soc.*, **2002**, *61*, 61-69.
- [78] Uauy, R.; Dangour, A.D. Nutrition in brain development and aging: role of essential fatty acids. *Nutr. Rev.*, **2006**, *64*, S24-33.

- [79] Haggarty, P. Placental regulation of fatty acid delivery and its effect on fetal growth--a review. *Placenta*, **2002**, *23*, S28-38.
- [80] Haggarty, P. Effect of placental function on fatty acid requirements during pregnancy. *Eur. J. Clin. Nutr.*, **2004**, *58*, 1559-1570.
- [81] Vancassel, S.; Durand, G.; C Barthelemy, C.; Lejeune, B.; Martineau, J.; Guilloteau, D.; Andres, C.; Chalon, S. Plasma fatty acid levels in autistic children. *Prostaglandins Leukot. Essent. Fatty Acids*, **2001**, *65*, 1-7.
- [82] Richardson, A. J.; Ross, M. A. Fatty acid metabolism in neurodevelopmental disorder: a new perspective on associations between attention-deficit/hyperactivity disorder, dyslexia, dyspraxia and the autistic spectrum. *Prostaglandins Leukot. Essent. Fatty Acids*, **2000**, *63*, 1-9.
- [83] Maloney, E. K.; Waxman, D. J. trans-Activation of PPARalpha and PPARgamma by structurally diverse environmental chemicals. *Toxicol. Appl. Pharmacol.*, **1999**, *161*, 209-218.
- [84] Xu, Y.; Knipp, G. T.; Cook, T. J. Effects of di-(2-ethylhexyl)phthalate and its metabolites on the lipid profiling in rat HRP-1 trophoblast cells. *Arch. Toxicol. Sci.*, **2006**, *80*, 293-298.
- [85] Moran, F. M.; Hendrickx, A. G.; Shideler, S.; Overstreet, J.W.; Watkins, S. M.; Lasley, B. L. Effects of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) on fatty acid availability and neural tube formation in cynomolgus macaque, *Macaca fascicularis*. *Birth Defects Res. B Dev. Reprod. Toxicol.*, **2004**, *71*, 37-46.
- [86] Shiota, K.; Mima, S. Assessment of the teratogenicity of di(2-ethylhexyl)phthalate and mono(2-ethylhexyl)phthalate in mice. *Arch. Toxicol.*, **1985**, *56*, 263-266.
- [87] Shea, K. M. Pediatric exposure and potential toxicity of phthalate plasticizers. *Pediatrics*, **2003**, *111*, 1467-1474.
- [88] Shiota, K.; Nishimura, H. Teratogenicity of di(2-ethylhexyl)phthalate (DEHP) and di-n-butyl phthalate (DBP) in mice. *Environ. Health Perspect.*, **1982**, *45*, 65-70.
- [89] Shiota, K.; Chou, M. J.; Nishimura, H. Embryotoxic effects of di-2-ethylhexyl phthalate (DEHP) and di-n-butyl phthalate (DBP) in mice. *Environ. Res.*, **1980**, *22*, 245-253.
- [90] Tanaka, J.; Yonemoto, J.; Zaha, H.; Kiyama, R.; Sone, H. Estrogen-responsive genes newly found to be modified by TCDD exposure in human cell lines and mouse systems. *Mol. Cell. Endocrinol.*, **2007**, *272*, 38-49.
- [91] Valdez, K. E.; Shi, Z.; Ting, A. Y.; Petroff, B. K. Effect of chronic exposure to the aryl hydrocarbon receptor agonist 2,3,7,8-tetrachlorodibenzo-p-dioxin in female rats on ovarian gene expression. *Reprod. Toxicol.*, **2009**, *28*, 32-37.
- [92] Skakkebaek, N.E.; Rajpert-De Meyts, E.; Main, K.M. Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects. *Hum. Reprod.*, **2001**, *16*, 972-928.
- [93] Anway, M.D.; Cupp, A.S.; Uzumcu, M.; Skinner, M.K. Epigenetic transgenerational actions of endocrine disruptors and male fertility. *Science*, **2005**, *308*, 1466-1469.
- [94] Main, K.M.; Skakkebaek, N.E.; Toppari, J. Cryptorchidism as part of the testicular dysgenesis syndrome: the environmental connection. *Endocr. Dev.*, **2009**, *14*, 167-173.
- [95] Damgaard, I.N.; Skakkebaek, N.E.; Toppari, J.; Virtanen, H.E.; Shen, H.; Schramm, K.W.; Petersen, J.H.; Jensen, T.K.; Main, K.M.; Nordic Cryptorchidism Study Group. Persistent pesticides in human breast milk and cryptorchidism. *Environ. Health Perspect.*, **2006**, *114*, 1133-1138.
- [96] Hosie, S.; Loff, S.; Witt, K.; Niessen, K.; Waag, K.L. Is there a correlation between organochlorine compounds and undescended testes? *Eur. J. Pediatr. Surg.*, **2000**, *10*, 304-309.
- [97] Langer, P.; Kocan, A.; Tajtáková, M.; Petřík, J.; Chovancová, J.; Drobná, B.; Jursa, S.; Pavúk, M.; Koska, J.; Trnovec, T.; Seböková, E.; Klimes, I. Possible effects of polychlorinated biphenyls and organochlorinated pesticides on the thyroid after long-term exposure to heavy environmental pollution. *J. Occup. Environ. Med.*, **2003**, *45*, 526-532.
- [98] Toppari, J.; Kaleva, M.; Virtanen, H.E. Trends in the incidence of cryptorchidism and hypospadias, and methodological limitations of registry-based data. *Hum. Reprod. Update*, **2001**, *7*, 282-286.
- [99] Pierik, F.H.; Burdorf, A.; Nijman, J.; de Muinck Keizer-Schrama, S.M.; Juttman, R.E.; Weber, R.F. A high hypospadias rate in The Netherlands. *Hum. Reprod.*, **2002**, *17*, 1112-1115.
- [100] Teilmann, G.; Juul, A.; Skakkebaek, N.E.; Toppari, J. Putative effects of endocrine disruptors on pubertal development in the human. *Best Pract. Res. Clin. Endocrinol. Metab.*, **2002**, *16*, 105-121.
- [101] Parent, A.S.; Teilmann, G.; Juul, A.; Skakkebaek, N.E.; Toppari, J.; Bourguignon, J.P. The timing of normal puberty and the age limits of sexual precocity: variations around the world, secular trends, and changes after migration. *Endocr. Rev.*, **2003**, *24*, 668-693.
- [102] Aksglaede, L.; Sørensen, K.; Petersen, J.H.; Skakkebaek, N.E.; Juul, A. Recent decline in age at breast development: the Copenhagen Puberty Study. *Pediatrics*, **2009**, *123*, e932-939.
- [103] Köhrle, J. Environment and endocrinology: the case of thyroidology. *Ann. Endocrinol. (Paris)*, **2008**, *69*, 116-122.
- [104] Koopman-Esseboom, C.; Morse, D.C.; Weisglas-Kuperus, N.; Lutkeschipholt, I.J.; Van der Paauw, C.G.; Tuinstra, L.G.; Brouwer, A.; Sauer, P.J. Effects of dioxins and polychlorinated biphenyls on thyroid hormone status of pregnant women and their infants. *Pediatr. Res.*, **1994**, *36*, 468-473.
- [105] Hsu, P.C.; Lai, T.J.; Guo, N.W.; Lambert, G.H.; Leon Guo, Y. Serum hormones in boys prenatally exposed to polychlorinated biphenyls and dibenzofurans. *J. Toxicol. Environ. Health A*, **2005**, *68*, 1447-1456.
- [106] Hagmar, L.; Rylander, L.; Dyremark, E.; Klasson-Wehler, E.; Erfurth, E.M. Plasma concentrations of persistent organochlorines in relation to thyrotropin and thyroid hormone levels in women. *Int. Arch. Occup. Environ. Health*, **2001**, *74*, 184-188.
- [107] Takser, L.; Mergler, D.; Baldwin, M.; de Grosbois, S.; Smargiassi, A.; Lafond, J. Thyroid hormones in pregnancy in relation to environmental exposure to organochlorine compounds and mercury. *Environ. Health Perspect.*, **2005**, *113*, 1039-1045.
- [108] Langer, P.; Kocan, A.; Tajtáková, M.; Petřík, J.; Chovancová, J.; Drobná, B.; Jursa, S.; Pavúk, M.; Koska, J.; Trnovec, T.; Seböková, E.; Klimes, I. Possible effects of polychlorinated biphenyls and organochlorinated pesticides on the thyroid after long-term exposure to heavy environmental pollution. *J. Occup. Environ. Med.*, **2003**, *45*, 526-532.
- [109] Sandau, C.D.; Ayotte, P.; Dewailly, E.; Duffe, J.; Norstrom, R.J. Pentachlorophenol and hydroxylated polychlorinated biphenyl metabolites in umbilical cord plasma of neonates from coastal populations in Québec. *Environ. Health Perspect.*, **2002**, *110*, 411-417.
- [110] Lindhe, O.; Lund, B.O.; Bergman, A.; Brandt, I. Irreversible binding and adrenocorticolytic activity of the DDT metabolite 3-methylsulfonyl-DDE examined in tissue-slice culture. *Environ. Health Perspect.*, **2001**, *109*, 105-110.
- [111] Anderson M.S. Update in endocrine autoimmunity. *J. Clin. Endocrinol. Metab.*, **2008**, *93*, 3663-70.
- [112] Owen C.J.; Cheetham T.D. Diagnosis and management of polyendocrinopathy syndromes. *Endocrinol. Metab. Clin. North. Am.*, **2009**, *38*, 419-436.
- [113] Ahmed S.A.; Talal N. Sex hormones and the immune system--Part 2. Animal data. *Baillieres Clin. Rheumatol.*, **1990**, *4*, 13-31.
- [114] Peeva E.; Zouali M. Spotlight on the role of hormonal factors in the emergence of autoreactive B-lymphocytes. *Immunol. Lett.*, **2005**, *101*, 123-143.
- [115] Ahmed S.A.; Hissong B.D.; Verthelyi D.; Donner K.; Becker K.; Karpuzoglu-Sahin E. Gender and risk of autoimmune diseases: possible role of estrogenic compounds. *Environ. Health Perspect.*, **1999**, *107*(Suppl 5), 681-686.
- [116] Zandman-Goddard G.; Peeva E.; Shoenfeld Y. Gender and autoimmunity. *Autoimmun. Rev.*, **2007**, *6*, 366-372.
- [117] Ahmed S.A. The immune system as a potential target for environmental estrogens (endocrine disruptors): a new emerging field. *Toxicology*, **2000**, *150*, 191-206.
- [118] Rao T.; Richardson B. Environmentally induced autoimmune diseases: potential mechanisms. *Environ. Health Perspect.*, **1999**, *107* Suppl 5, 737-742.
- [119] Yurino H, Ishikawa S, Sato T, Akadegawa K, Ito T, Ueha S, Inadera H, Matsushima K. Endocrine disruptors (environmental estrogens) enhance autoantibody production by B1 cells. *Toxicol. Sci.*, **2004**, *81*, 139-147
- [120] Guarneri F.; Benvenga S. Environmental factors and genetic background that interact to cause autoimmune thyroid disease. *Curr. Opin. Endocrinol. Diabetes Obes.*, **2007**, *14*, 398-409.
- [121] Ablake, M.; Itoh, M.; Terayama, H.; Hayashi, S.; Shoji, S.; Naito, M.; Takahashi, K.; Suna, S.; Jitsunari, F. Di-(2-ethylhexyl) phthalate induces severe aspermatogenesis in mice, however, subsequent

- antioxidant vitamins supplementation accelerates regeneration of the seminiferous epithelium. *Int. J. Androl.*, **2004**, *27*, 274-281.
- [122] Albertini, R.; Bird, M.; Doerrer, N.; Needham, L.; Robison, S.; Sheldon, L.; Zenick, H. The use of biomonitoring data in exposure and human health risk assessments. *Environ. Health Perspect.*, **2006**, *114*, 1755-1762.
- [123] Angerer, J.; Ewers, U.; Wilhelm, M. Human biomonitoring: state of the art. *Int. J. Hyg. Environ. Health*, **2007**, *210*, 201-228.
- [124] Wormuth, M.; Demou, E.; Scheringer, M.; Hungerbühler, K. Assessments of direct human exposure: the approach of EU risk assessments compared to scenario-based risk assessment. *Risk Anal.*, **2007**, *27*, 979-990.
- [125] Hines, E.P.; Calafat, A.M.; Silva, M.J.; Mendola, P.; Fenton, S.E. Concentrations of phthalate metabolites in milk, urine, saliva, and Serum of lactating North Carolina women. *Environ. Health Perspect.*, **2009**, *117*, 86-92.
- [126] Latini, G.; Wittassek, M.; Del Vecchio, A.; Presta, G.; De Felice, C.; Angerer, J. Lactational exposure to phthalates in Southern Italy. *Environ. Int.*, **2009**, *35*, 236-239.
- [127] Mortensen, G.K.; Main, K.M.; Andersson, A.M.; Leffers, H.; Skakkebaek, N.E. Determination of phthalate monoesters in human milk, consumer milk, and infant formula by tandem mass spectrometry (LC-MS-MS). *Anal. Bioanal. Chem.*, **2005**, *382*, 1084-1092.
- [128] Fromme, H.; Gruber, L.; Schlummer, M.; Wolz, G.; Böhmer, S.; Angerer, J.; Mayer, R.; Liebl, B.; Bolte, G. Intake of phthalates and di(2-ethylhexyl)adipate: results of the Integrated Exposure Assessment Survey based on duplicate diet samples and biomonitoring data. *Environ. Int.*, **2007**, *33*, 1012-1020.
- [129] Seckin, E.; Fromme, H.; Völkel, W. Determination of total and free mono-n-butyl phthalate in human urine samples after medication of a di-n-butyl phthalate containing capsule. *Toxicol. Lett.*, **2009**, *188*, 33-37.
- [130] Wensing, M.; Uhde, E.; Salthammer, T. Plastics additives in the indoor environment--flame retardants and plasticizers. *Sci. Total Environ.*, **2005**, *339*, 19-40.
- [131] Koo, H.J.; Lee, B.M. Estimated exposure to phthalates in cosmetics and risk assessment. *J. Toxicol. Environ. Health A.*, **2004**, *67*, 1901-1914.
- [132] Latini, G. Monitoring phthalate exposure in humans. *Clin. Chim. Acta*, **2005**, *361*, 20-29.
- [133] Heudorf, U.; Mersch-Sundermann, V.; Angerer, J. Phthalates: toxicology and exposure. *Int. J. Hyg. Environ. Health.*, **2007**, *210*, 623-634.
- [134] Koch, H.M.; Rossbach, B.; Drexler, H.; Angerer, J. Internal exposure of the general population to DEHP and other phthalates--determination of secondary and primary phthalate monoester metabolites in urine. *Environ. Res.*, **2003**, *93*, 177-185.
- [135] Koch, H.M.; Drexler, H.; Angerer, J. Internal exposure of nursery-school children and their parents and teachers to di(2-ethylhexyl)phthalate (DEHP). *Int. J. Hyg. Environ. Health.*, **2004**, *207*, 15-22.
- [136] Calafat, A.M. McKee R.H. Integrating biomonitoring exposure data into the risk assessment process: phthalates [diethyl phthalate and di(2-ethylhexyl) phthalate] as a case study. *Environ. Health Perspect.*, **2006**, *114*, 1783-1789
- [137] Koo, J.W.; Parham, F.; Kohn, M.C.; Masten, S.A.; Brock, J.W.; Needham, L.L.; Portier, C.J. The association between biomarker-based exposure estimates for phthalates and demographic factors in a human reference population. *Environ Health Perspect.*, **2002**, *110*, 405-410.
- [138] Blount, B.C.; Milgram, K.E.; Silva, M.J.; Malek, N.A.; Reidy, J.A.; Needham, L.L.; Brock, J.W. Quantitative detection of eight phthalate metabolites in human urine using HPLC-APCI-MS/MS. *Anal. Chem.*, **2000**, *72*, 4127-4134.
- [139] Hatch, E.E.; Nelson, J.W.; Qureshi, M.M.; Weinberg, J.; Moore, L.L.; Singer, M.; Webster, T.F. Association of urinary phthalate metabolite concentrations with body mass index and waist circumference: a cross-sectional study of NHANES data, 1999-2002. *Environ. Health.*, **2008**, *7*, 27.
- [140] Latini, G.; De Felice, C.; Presta, G.; Del Vecchio, A.; Paris, I.; Ruggirei, F.; Mazzeo, P. In utero exposure to di(2-ethylhexyl)phthalate and duration of human pregnancy. *Environ. Health Perspect.*, **2003**, *111*, 1783-1785.
- [141] Meeker, J.D.; Calafat, A.M.; Hauser, R. Di(2-ethylhexyl) phthalate metabolites may alter thyroid hormone levels in men. *Environ. Health Perspect.*, **2007**, *115*, 1029-1034.
- [142] Hokanson, R.; Hanneman, W.; Hennessey, M.; Donnelly, K.C.; McDonald, T.; Chowdhary, R.; Busbee, D.L. DEHP, bis(2)-ethylhexyl phthalate, alters gene expression in human cells: possible correlation with initiation of fetal developmental abnormalities. *Hum. Exp. Toxicol.*, **2006**, *25*, 687-695.
- [143] Jaakkola, J.J.; Knight, T.L. The role of exposure to phthalates from polyvinyl chloride products in the development of asthma and allergies: a systematic review and meta-analysis. *Environ. Health Perspect.*, **2008**, *116*, 845-853.
- [144] Palleschi, S.; Rossi, B.; Diana, L.; Silvestroni, L. Di(2-ethylhexyl)phthalate stimulates Ca(2+) entry, chemotaxis and ROS production in human granulocytes. *Toxicol. Lett.*, **2009**, *187*, 52-57.
- [145] Wittassek, M.; Angerer, J.; Kolossa-Gehring, M.; Schäfer, S.D.; Klockenbusch, W.; Dobler, L.; Günzel, A.K.; Müller, A.; Wiesmüller, G.A. Fetal exposure to phthalates - a pilot study. *Int. J. Hyg. Environ. Health.*, **2009**, *212*(6), 685-92.
- [146] Fabjan, E.; Hulzebos, E.; Mennes, W.; Piersma, A.H. A category approach for reproductive effects of phthalates. *Crit. Rev. Toxicol.*, **2006**, *36*, 695-726.
- [147] Wirth, J.J.; Rossano, M.G.; Potter, R.; Puscheck, E.; Daly, D.C.; Paneth, N.; Krawetz, S.A.; Protas, B.M.; Diamond, M.P. A pilot study associating urinary concentrations of phthalate metabolites and semen quality. *Syst. Biol. Reprod. Med.*, **2008**, *54*, 143-154.
- [148] Swan, S.H. Environmental phthalate exposure in relation to reproductive outcomes and other health endpoints in humans. *Environ. Res.*, **2008**, *108*, 177-184.
- [149] Lovekamp-Swan, T.; Davis, B.J. Mechanisms of phthalate ester toxicity in the female reproductive system. *Environ. Health Perspect.*, **2003**, *111*, 139-145.
- [150] Latini, G.; Del Vecchio, A.; Massaro, M.; Verrotti, A.; De Felice, C. Phthalate exposure and male infertility. *Toxicology*, **2006**, *226*, 90-98.
- [151] McKee, R.H.; Butala, J.H.; David, R.M.; Gans, G. NTP center for the evaluation of risks to human reproduction reports on phthalates: addressing the data gaps. *Reprod. Toxicol.*, **2004**, *18*, 1-22.
- [152] Heudorf, U.; Mersch-Sundermann, V.; Angerer, J. Phthalates: toxicology and exposure. *Int. J. Hyg. Environ. Health*, **2007**, *210*, 623-634.
- [153] Center for Devices and Radiological Health, U.S. Food and Drug Administration. Safety Assessment of Di(2-ethylhexyl)phthalate (DEHP) Released from PVC Medical Devices. Available at <http://www.fda.gov/cdrh/ost/dehp-pvc.pdf>. [accessed 5 September 2001]
- [154] Koch, H.M.; Preuss R.; Angerer J. Di(2-ethylhexyl)phthalate (DEHP): human metabolism and internal exposure—an update and latest results. *Int. J. Androl.*, **2006**, *29*, 155-165.
- [155] Kamrin, M.A. Phthalate risks, phthalate regulation, and public health: a review. *J. Toxicol. Environ. Health B. Crit. Rev.*, **2009**, *12*, 157-174.
- [156] Wormuth, M.; Demou, E.; Scheringer, M.; Hungerbühler, K. Assessments of direct human exposure: the approach of EU risk assessments compared to scenario-based risk assessment. *Risk Anal.*, **2007**, *27*, 979-990.